## Temporal characteristic simulation for stimulated emission depletion microscopy

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The imaging technology of stimulated emission depletion (STED) utilizes the nonlinearity relationship between the fluorescence saturation and the excited state stimulated depletion. It implements three-dimensional (3D) imaging and breaks the diffraction barrier of far-field light microscopy by restricting fluorescent molecules at a sub-diffraction spot. In order to improve the resolution which attained by this technology, the computer simulation on temporal behavior of population probabilities of the sample was made in this paper, and the optimized parameters such as intensity, duration and delay time of the STED pulse were given.

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Far-field fluorescence light microscopy is a main technique for investigating and detecting biological specimens, because the focused beams can penetrate high scatter media with the intention of imaging living specimens with three-dimensional (3D). But its resolution is limited by the diffraction barrier<sup>[1]</sup> which depends on the smallest focusing light spot size.

The diffraction barrier and limited resolution in farfield fluorescence microscopy has been fundamentally broken by utilizing the nonlinearity relationship between the fluorescence saturation and the excited state stimulated depletion, which called stimulated emission depletion (STED). It is reported that Hell team have attained spots of 33-nm width along the optic axis [2] and lateral resolution of 28 nm, which corresponding to 1/25 of wavelength<sup>[3]</sup>. This improvement is obtained by quenching the excited fluorescence at the out part of the excitation focal spot with a beam whose wavelength is at the red edge of the fluorescent emission spectrum. Consequently the remaining fluorescence stems from a region that is narrower than the diffraction-limited size.

There are two kinds issues in the STED fluorescence microscopy, one is the spatial shaping of the stimulating beam, so as to render a focal spot with a central hole; the other is the spectroscopy and aims at realizing a set of conditions that enables the complete depletion of the excited state by stimulated emission.

In theory, the ideal focal intensity distribution of the STED pulse should subject to the function  $I_{\text{STED}}(\vec{r},t) = \left\{ \begin{array}{ll} 0, & \text{for } \vec{r}=0 \\ \gg 0, & \text{else} \end{array} \right.$ , which

ensures that the center of the fluorescence spot is spared out from depletion, whereas its periphery there the fluorescent molecules are strongly depleted. Therefore only molecules, which happen to be close to the focal point, contribute to the detected signal, so theoretically the achievable spatial resolution is on a molecular scale. But in practice it is limited by the finite depth of the central minimum formed by the molecules, and the resolution will be further increased if larger depletion efficiency can be attained. So the computer simulation on the temporal behaviors of population probabilities of the sample was made in this paper, and the optimized parameters such as intensity, duration, and delay time of the STED pulse

were given to improve the depletion efficiency.

Meeting the spectroscopic conditions is based on the fact that actually all fluorophores are four-level systems. Taking this into account, the energy levers involved in the excitation and the subsequent emission process of a typical fluorophore was displayed in Fig. 1<sup>[5]</sup>.

After its excitation to a higher vibrational level  $L_1$  of the first excited electronic state  $S_1$ , the molecules nonradiatively decay to the low-lying fluorescent state  $L_2$ . The red shifted depletion pulse induces stimulated emission in  $L_2$  and pushes the molecules to a higher vibrational level  $L_3$  of the ground state  $S_0$ , which decays to the low vibrational level  $L_0$ .

The temporal behaviors of the population probabilities  $n_i$  (t) of levels  $L_i$  (i = 0, 1, 2, 3) of the dye can be described by a set of coupled differential equations, relating the interplay among absorption, quenching, vibrational relaxation, stimulated emission, and spontaneous emission<sup>[4,5]</sup>:

$$\frac{dn_0}{dt} = \frac{I_{\text{exc}}\sigma_{01}(n_1 - n_0)}{h\nu_{\text{exc}}} + \frac{1}{\tau_{\text{vibr}}}n_3,$$
(1)

$$\frac{dn_1}{dt} = \frac{I_{\text{exc}}\sigma_{01}(n_0 - n_1)}{h\nu_{\text{exc}}} - \frac{1}{\tau_{\text{vibr}}} n_1,$$
 (2)

$$\frac{\mathrm{d}n_2}{\mathrm{d}t} = \frac{I_{\mathrm{STED}}\sigma_{23}(n_3 - n_2)}{h\nu_{\mathrm{STED}}} + \frac{1}{\tau_{\mathrm{vibr}}} n_1 - (\frac{1}{\tau_{\mathrm{fluor}}} + Q)n_2, \tag{3}$$

$$\frac{\mathrm{d}n_3}{\mathrm{d}t} = \frac{I_{\mathrm{STED}}\sigma_{23}(n_2 - n_3)}{h\nu_{\mathrm{STED}}} - \frac{1}{\tau_{\mathrm{vibr}}} n_3 + (\frac{1}{\tau_{\mathrm{fluor}}} + Q)n_2, \tag{4}$$

$$\sum_{i} n_i = 1, \tag{5}$$

$$n_0(t=0) = 1, (6)$$

where Q is the quenching rate,  $\tau_{\rm fluor}$  and  $\tau_{\rm vibr}$  are the average fluorescence lifetime and vibrational relaxation

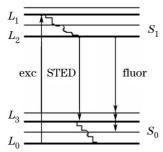


Fig. 1. Energy levels of a typical fluorophore.

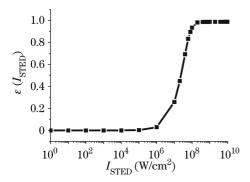


Fig. 2. Time-averaged depletion efficiency as a function of  $I_{\text{STED}}$ .

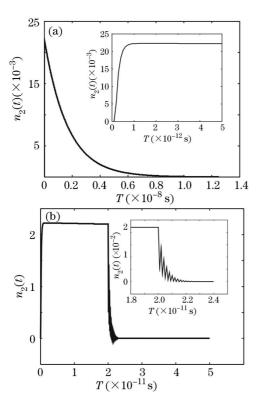


Fig. 3. Temporal behaviors of population probabilities  $n_2$ , with  $I_{\rm STED}=0(a)$  and  $I_{\rm STED}=10^{10}$  W/cm<sup>2</sup> (b).

time;  $\sigma_{01}$  and  $\sigma_{23}$  are the molecular cross-section for the transition  $L_0 \leftrightarrow L_1$  and  $L_2 \leftrightarrow L_3$ ;  $h\nu_{\rm exc}$  and  $h\nu_{\rm STED}$  are the photon energies and  $I_{\rm exc}$  and  $I_{\rm STED}$  are the local intensities of excitation and STED beams, respectively.  $\frac{I_{\rm exc}\,\sigma_{01}(n_1-n_0)}{h\nu_{\rm exc}} \ {\rm describes} \ {\rm the} \ {\rm absorption} \ {\rm and} \ {\rm dem} \ {\rm intensition} \ {\rm the} \ {\rm t$ 

the stimulated emission from  $L_2$  and re-excitation from  $L_3$  by stimulating beam, and  $(\frac{1}{\tau_{\text{fluor}}} + Q)n_2$  describes the decay of  $n_2$  by spontaneous emission and quenching. Equations (5) and (6) denote the normalized probabilities for the total population and the initialization.

To simulate the temporal behaviors of  $n_i$ , some parameters of the coupled differential equations are set as follow:  $\sigma_{01} = 4 \times 10^{-16} \text{ cm}^2$ ,  $\sigma_{23} = 2 \times 10^{-16} \text{ cm}^2$ ,  $\tau_{\text{fluor}} = 2 \times 10^{-9} \text{ s}$ ,  $\tau_{\text{vibr}} = 0.2 \times 10^{-12} \text{ s}$ ,  $Q = 10^8 \text{ s}^{-1}$ ,  $\lambda_{\text{exc}} = 560 \times 10^{-9} \text{ m}$ ,  $\lambda_{\text{exc}} = 765 \times 10^{-9} \text{ m}$ . Here the fluorescence wavelengths range from 650 to 800 nm<sup>[6]</sup>. The excitation and STED pulses are supposed as rectangular pulses and their mathematic expressions are given

gular pulses and their mathematic expressions are given as 
$$I_{\rm exc}(t) = \begin{cases} I_0^{\rm exc}, & {\rm for} 0 \leq t \leq \tau_{\rm exc} \\ 0, & {\rm else} \end{cases}$$
 and  $I_{\rm STED}(t) = \begin{cases} I_0^{\rm STED} \cdots {\rm for} \cdots 0 \leq t \leq \tau_{\rm STED} \\ 0 \cdots \cdots {\rm else} \end{cases}$ ,  $\tau_{\rm exc}$  and  $\tau_{\rm STED}$  refer to the duration time of excitation and STED pulses, re-

spectively. Delay time  $\Delta t$  denotes the time between the climbing-edge of excitation and STED pulses.

Only to demonstrate that the temporal behaviors of  $n_i$  is not enough, so we also simulate the time-averaged depletion efficiency  $\varepsilon$  of the fluorescent state, which is

defined as 
$$\varepsilon = \varepsilon(I_{\rm STED}) = 1 - \frac{\int\limits_0^T n_2(t,I_{\rm STED}(t))\,\mathrm{d}t}{\int\limits_0^T n_2(t,0)\,\mathrm{d}t}$$
,  $T$  is the

integral time, which is set as the interval time of pluses train here.

Firstly, the nonlinear relationship between STED intensity and time-averaged depletion efficiency of the fluorescent state are demonstrated in Fig. 2.  $I_{\text{STED}}$  varies from 0 to  $10^{10} \mathrm{\ W/cm}^2$  and other parameters are selected as follow:  $I_{\rm exc} = 10^8 \ {\rm W/cm^2}, \ \tau_{\rm exc} = 0.2 \times 10^{-12} \ {\rm s}, \ \tau_{\rm STED} = 40 \times 10^{-12} \ {\rm s}, \ \Delta t = 20 \times 10^{-12} \ {\rm s}.$  Figure 2 shows that when  $I_{\rm STED} \leq 10^6 {\rm W/cm^2}$ , the time-averaged depletion efficiency  $\varepsilon$  is almost not change and approximates to 0; when  $10^6 \le I_{\text{STED}} \le 2 \times 10^8 \text{ W/cm}^2$ , with intensity increasing,  $\varepsilon$  increases sharply, and changes from 0.03 to 0.9820; with  $I_{\rm STED}$  continuing increasing,  $\varepsilon$  is approaching saturation and close to 100%. We can conclude that when  $I_{\rm STED} \leq 10^6 \ {\rm W/cm^2}$  which may be named as "threshold intensity", or  $I_{\text{STED}} \geq 2 \times 10^8 \text{ W/cm}^2$ , the temporal behaviors of the population probabilities  $n_2$  are almost of the same, respectively. Contrastively, when  $I_{\rm STED}$  at the range of  $10^6 \le I_{\rm STED} \le 2 \times 10^8$  $W/cm^2$ , the temporal behaviors of  $n_2$  depend on  $I_{STED}$ acutely. Consequent we will analysis the temporal behaviors of  $n_2$  at different typical intensities.

Observing Fig. 3, at the initial time, the fluorescent state is empty. After 0.2 ps, particles begin to occupy  $L_2$  and the population adds, at about 1 ps the population reaches its maximum. Then if  $I_{\rm STED}$  is low enough as to render the rates of the stimulated transition much slower than  $1/\tau_{\rm vibr}$ , re-excitation is negligible, and the vibrational state near empty, so  $n_1 \approx 0$ ,  $n_3 \approx 0$ . Hence, the population of the fluorescent state and the remaining fluorescence obeys the proportionality  $n_2(t) \propto \exp[-(\frac{1}{\tau_{\rm fluor}} + Q)t]$  in good approximation, this means that  $n_2$  is governed by the fluorescence emission rate. But for higher intensity, after the population reached the maximum, it keeps constant during the

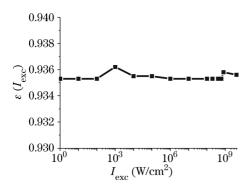


Fig. 4. Time-averaged depletion efficiency as a function of  $I_{\rm exc}$ .

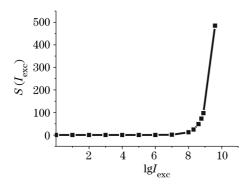


Fig. 5. The sum of  $n_2$  as a function the excitation intensity  $I_{\rm exc}$ .

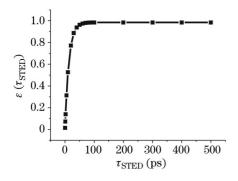


Fig. 6. Time-averaged depletion efficiency as a function of  $\tau_{\text{STED}}$ .

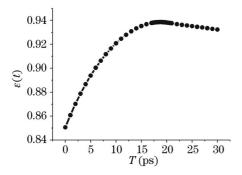


Fig. 7. Time-averaged depletion efficiency as a function of the delay time  $\Delta t$ .

delay time, when STED pulse begins to operate,  $n_2$  decays faster than the low intensity case, and as the whole

procession it is determined by the fast vibrational relaxation but with damping oscillation. For the condition that  $10^6 \leq I_{\rm STED} \leq 2 \times 10^8 \ {\rm W/cm^2}$ , the temporal behavior of  $n_2$  depends on  $I_{\rm STED}$  and the decay time changes from  $\tau_{\rm fluor}$  to  $\tau_{\rm vibr}$ , because all influence of fluorescence emission, quenching, STED intensity, and vibrational relaxation cannot be neglected. Moreover, since  $\sigma_{23}$  plays the same role of  $I_{\rm STED}$  in Eq. (3), we can select samples with big  $\sigma_{23}$  to enhance the depletion efficiency.

Then the influence of  $I_{\rm exc}$  to the depletion efficiency is considered. We Keep the intensity  $I_{\rm STED}=10^8~{\rm W/cm^2}$ , and change the value of  $I_{\rm exc}$  from zero to  $10^{10}~{\rm W/cm^2}$ . In Fig. 4, we can see that  $\varepsilon$  approximately keeps the same value. But when consider the sum of population probabilities of fluorescent state in a period interval which de-

fined as  $S(I_{\rm exc}) = \int\limits_0^T n_2(t,I_{\rm exc}) {\rm d}t$ . In Fig. 5, we find that with  $I_{\rm exc}$  increasing, the total signal  $S(I_{\rm exc})$  increases by degrees, but when  $I_{\rm exc} \leq 10^7 \ {\rm W/cm^2}$ , it nears zero and are too small to demonstrate its different values in the picture, if  $I_{\rm exc} \geq 10^7 \ {\rm W/cm^2}$ , it adds sharply. Therefore, though  $I_{\rm exc}$  almost does not influnce the depletion efficiency, while considering about other background light in the experiment, in order to improve the signal-to-noise ratio, the detected signal is better to be bigger, so we had better to set  $I_{\rm exc}$  with bigger value.

When investigate the duration of excitation pulse, we get the conclusion that increasing the pulse duration is equal to increasing the intensity, because increasing either the duration or intensity, it all adds the population of the photons per pulse, and it means more photons are pumped then down to the fluorescent state.

Next we let  $\tau_{\rm STED}$  increases from 0.2 to 500 ps, and keep  $I_{\rm STED}=10^8~{\rm W/cm^2}$ , change  $\tau_{\rm exc}$  into 20 ps to distinguish the influence of  $\tau_{\rm exc}$  and  $\tau_{\rm vibr}$ . The numerical results are described as the curve of the time-averaged depletion efficiency as a function of  $\tau_{\rm STED}$  in Fig 6. In which the curve climbs fast at the beginning, and then rise slowly, when  $\tau_{\rm STED} \geq 200$  ps, it becomes a constant. Comparing Figs.6 and 2, the two curves almost move in the same rule, that means that increasing the duration is equal to increasing the intensity, for the reason that they all adds the population of the photons per pulse.

Since the typical life time of the fluorescent state is about 1–5 ns, STED pulse has to be completed within a fraction of this time. This condition gives an upper limit of approximately 200–1000 ps for the duration of STED pulse. Conversely, there is a lower limit of approximately 1–2 ps which stems from the fact that transient filling of the higher vibrational level of the ground state must be avoided<sup>[7]</sup>.

Then we study the key parameter: the delay time  $\Delta t$ , which refers to the time that STED pulse after the excitation pulse, the accurate definition is given above. We set  $\tau_{\rm exc} = 20 \times 10^{-12} \, {\rm s}$ ,  $\tau_{\rm STED} = 40 \times 10^{-12} \, {\rm s}$ , others keep the same values, and add  $\Delta t$  from 0 to 30 ps step by step, around the maximum, we insert some points with 0.2-ps interval which equal to  $\tau_{\rm vibr}$  to see about whether  $\tau_{\rm vibr}$  has some infection on the depletion efficiency.

From Fig. 7, we can see that with the increasing of the delay time,  $\varepsilon$  becomes bigger, when 17.4 (ps)  $\leq \Delta t \leq 20.2$  (ps) which is be close to the duration of the excitation

pulse,  $\varepsilon$  is larger than 0.938, and at 18.8 ps , it gets to the maximum, then with bigger delay time, it declines slowly. We can conclude that the optimal value of  $\Delta t$  is such that the STED pulse arrives as soon as the excitation pulse is about to leave, which can be expressed as  $\tau_{\rm exc}-2.6$  (ps)  $\leq \Delta t_{\rm opt} \leq \tau_{\rm exc}+0.2$  (ps). In this case, within the restricted sub-diffraction spot,  $L_2$  is almost not being populated while the fluorescence is radiating.

In conclusion, we have investigated the temporal behaviors of the population probabilities of the excited level of a material, and the relationships between the time-averaged depletion efficiency and the key parameters such as  $I_{\text{exc}}$ ,  $\tau_{\text{exc}}$ ,  $I_{\text{STED}}$ ,  $\tau_{\text{STED}}$ , and  $\Delta t$ . We gain the rules as following: first, increasing  $I_{\text{exc}}$  and  $\tau_{\rm ex\,c}$  are equal to adds the photons per pulse of excitation beam, although they not to enhance efficiency directly, they increase the total signal; second, increasing  $I_{\rm STED}$  and  $au_{\rm STED}$  are equal to adds the photons per pulse of STED beam, and before their saturation, the bigger they are the higher efficiency it gets; third, when  $\tau_{\rm exc} - 2.6 \; (\rm ps) \leq \Delta t_{\rm opt} \leq \tau_{\rm exc} + 0.2 \; (\rm ps), \; STED \; light$ beam just reaches as soon as the excitation pulse is about to leave, it gets the maximum efficiency. In STED microscopy, the time size control is vital technique for improving the system resolution.

Through above simulation calculation and analysis, we expect the resolution can reach the molecule scale, and

is able to image inside a 3D volume, due to its sectional function, and it could be implemented in standard cofocal beam scanners, also it can be combined with total internal reflection microscopy to create spatially controlled excitation profiles<sup>[8]</sup>. STED microscopy should play a more and more important role in the future.

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